

Validation of the Applied Biosystems® 3500 Genetic Analyzer with a comparison of the Identifiler® Plus and PowerPlex® 16 HS amplification kits Emilie Dembia, B.S.¹; Sarah Chenoweth, M.S.²; Jennifer Hayden, M.S.¹; Pamela J. Staton, Ph.D.¹

Abstract

Anne Arundel County Crime Laboratory upgraded from an Applied Biosystems® (AB) 310 genetic analyzer to an AB 3500 genetic analyzer. The internal validation of this AB 3500 included a comparison of the PowerPlex® 16 HS (PP16HS) and Identifiler[®] Plus (ID+) amplification kits using the manufacturer's recommended protocols to determine if one kit had any benefit to forensic casework analysis over the other, when used in conjunction with the AB 3500.

Introduction

Based on a previous validation of the AB 3500 by the Mansfield Police Laboratory, injection times of 7s and 15s at 1.2kV were validated at the Anne Arundel County Crime Laboratory. Forensic casework samples were used to perform the following studies:

- •Analytical Thresholds* •Contamination* •Stochastic Thresholds •Concordance •Denaturation-Snap •Reproducibility* Cooling* •Stutter
- •Sensitivity*

•Precision*

- •Consumables Study
- •PP16HS vs. ID+*

Duplicate amplifications were performed with PP16HS and ID+ for the studies listed with an (*) for amplification kit comparison.

Materials and Methods

Instrumentation: •Qiagen BioRobot EZ1 Advanced XL - Trace TD protocol, elution in water •AB 7500 Real-Time PCR System •PowerPlex® 16 HS Amplification System •AmpFlSTR® Identifiler® Plus Amplification Kit •AB GeneAmp® PCR System 9700 •AB 3500 Genetic Analyzer Samples: •NIST Standard Reference Materials 11 and 12 •9947A Dilution Series - 0.1ng, 0.25ng, 0.5ng, 0.75ng, 1.0ng, 1.5ng, 2.5ng •Known male and known female 1:1 mixtures - 0.2ng, 0.6ng, 1.0ng •Known heterozygous individual's DNA extract •96-well checkerboards of allelic ladders and run negatives •24 ID+ re-amplifications of PP16HS casework samples •57 PP16HS casework samples at 0.6ng or 0.7ng •18 PP16HS casework samples re-amplified at 1ng **Analysis Software:**

•GeneMapper® ID-X version 1.2

PP16HS

Dye C

B

Gr

Yel Univ

ID+7sDye C

Univ

PP16HS and ID+ Comparison Study

| Percent Dropout | 80% |
|-----------------|-----|
| | 70% |
| | 60% |
| | 50% |
| | 40% |
| | 30% |
| | 20% |
| | 10% |
| | 0% |
| | |

Figure 1: Less dropout is observed with PP16HS compared to ID+ when using manufacturer's recommended protocols. Mixtures show the same results.

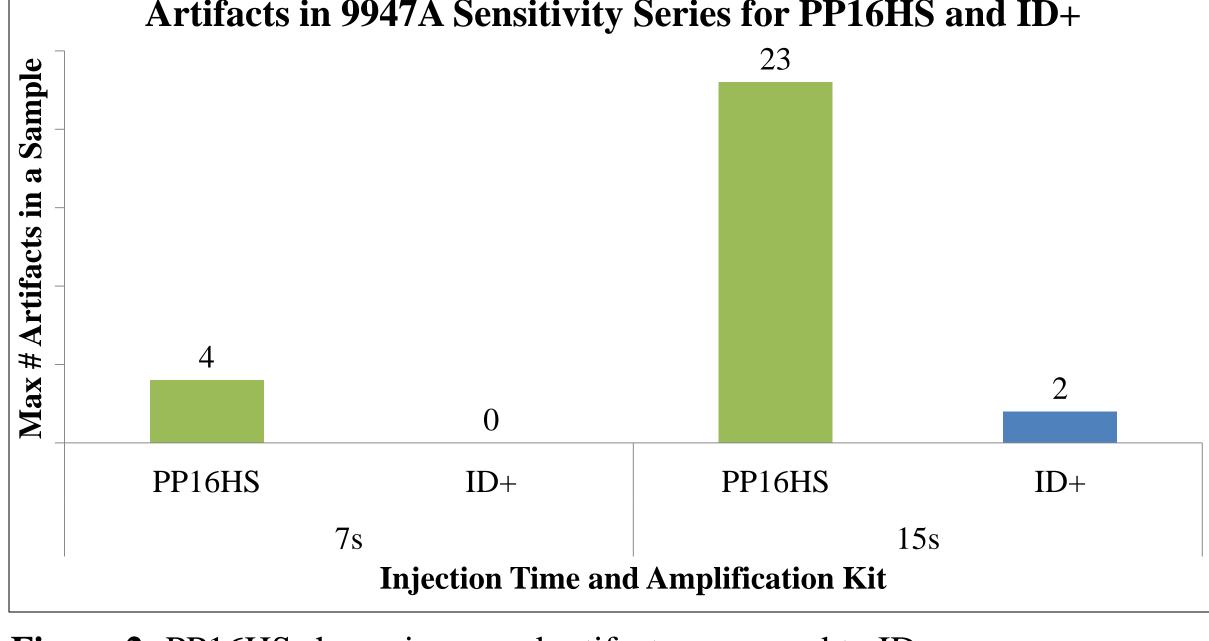


Figure 2: PP16HS shows increased artifacts compared to ID+.

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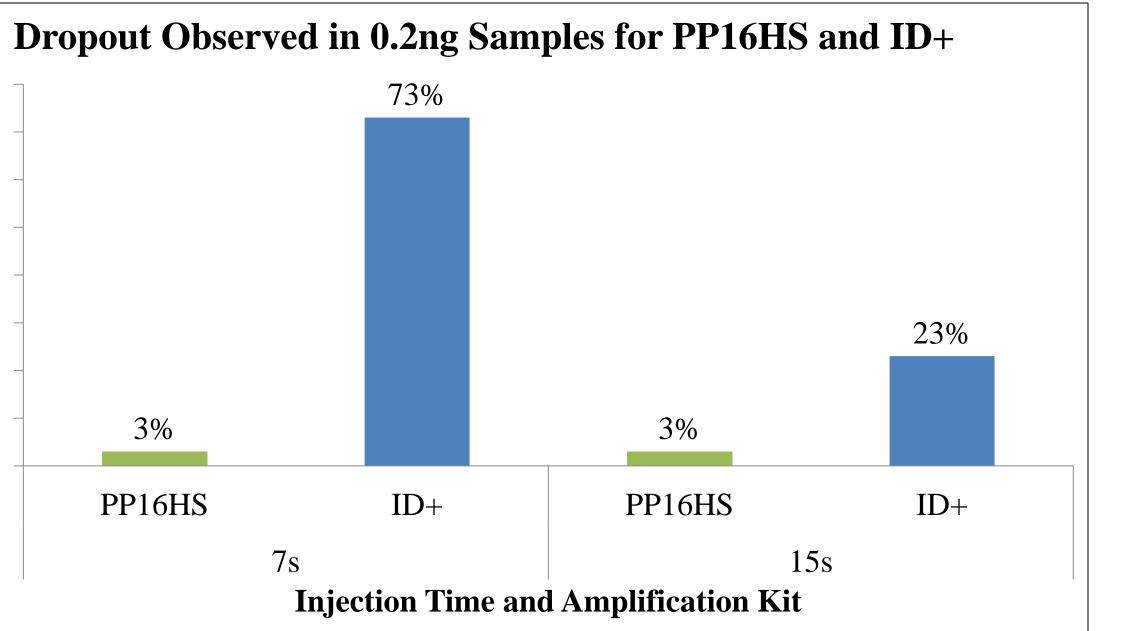
Results

Analytical Threshold Study

Table 1: Dye specific analytical thresholds were calculated, but universal thresholds were chosen for each chemistry at each injection time to give a more conservative value.

| S 7s Analytical Thresholds PP16HS 15s Analytical Thresholds | | | |
|---|-------------------------|-------------|-------------------------|
| Channel | Analytical Threshold | Dye Channel | Analytical Threshold |
| Blue | 45 RFU | Blue | 55 RFU |
| reen | 50 RFU | Green | 85 RFU |
| ellow | 55 RFU | Yellow | 100 RFU |
| versal | 75 RFU | Universal | 100 RFU |

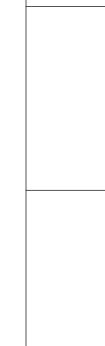
| - 7s Analytical Thresholds | | ID+15s Analytical Thresholds | |
|----------------------------|-------------------------|------------------------------|-------------------------|
| ye Channel | Analytical Threshold | Dye Channel | Analytical Threshold |
| Blue | 25 RFU | Blue | 25 RFU |
| Green | 40 RFU | Green | 45 RFU |
| Yellow | 70 RFU | Yellow | 80 RFU |
| Red | 100 RFU | Red | 110 RFU |
| Universal | 100 RFU | Universal | 125 RFU |



Artifacts in 9947A Sensitivity Series for PP16HS and ID+



Sensitivity Study Table 2: Lowest amount of DNA (9947A) to yield a full profile on the AB 3500.



| Injection Time | Amplification Kit | 1 st Complete DNA Profile |
|-------------------|----------------------|---|
| 7s | PP16HS | ≤50pg / 50pg |
| | ID+ | 50pg |
| 15s | PP16HS | ≤50pg /≤50pg |
| | ID+ | 50pg |

Table 3: Lowest amount of DNA (9947A) to show good peak height ratios (>50%) on the AB 3500. 7 randomly chosen single source samples showed an average peak height ratio of 0.83 for both PP16HS and ID+.

| Injection Time | Amplification Kit | Amplified DNA | Lowest PHR |
|-------------------|----------------------|------------------|---------------|
| 7s | PP16HS | ≤50pg | 55% |
| 7.5 | ID+ | 90pg | 53% |
| 15s | PP16HS | 50pg | 60% |
| | ID+ | 90pg | 54% |

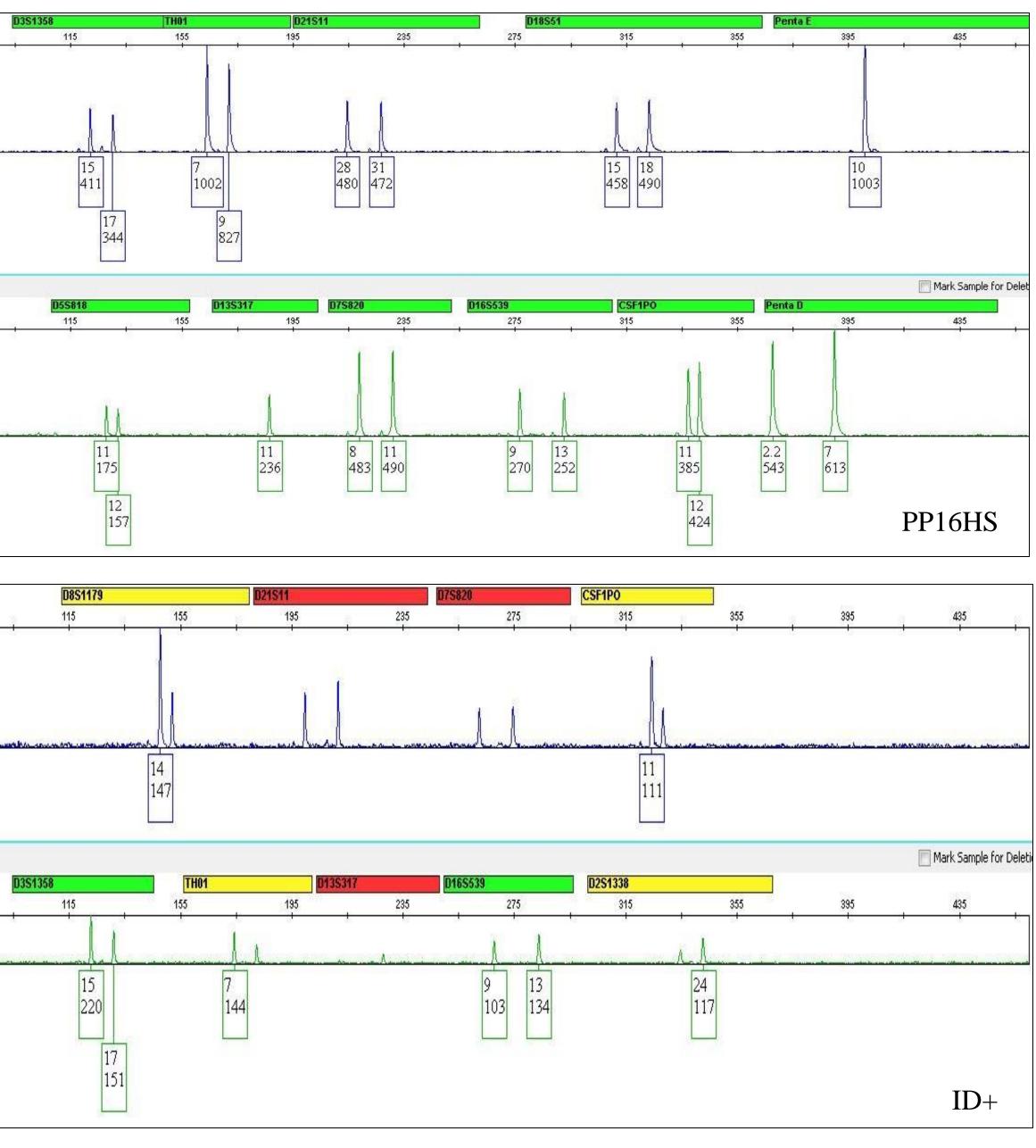


Figure 3: Non-probative casework sample CS-1-A amplified at 0.7ng with PP16HS (top) and ID+ (bottom). PP16HS recovered a full profile and ID+ recovered a partial profile using the 7s injection time on the AB 3500. Blue and green dye channels are shown for each amplification kit.



Discussion and Conclusions

•Amplification target: 0.6ng to 1.0ng DNA

•Stochastic Thresholds for PP16HS: 7s - 425 RFU 15s - 600 RFU

•Precision:

PP16HS kit standard deviation – 0.046 ID+ kit standard deviation -0.058

•Concordance: average decrease of ~210 RFU from AB 310 to AB 3500. All allele calls remained constant.

• Denaturation and snap cooling prior to an instrument run does not improve data produced by the AB 3500.

•POP 4 has a recommended usage time of 7 days after installation, but remains usable for an extended period if kept in the fridge when not on the instrument.

• PP16HS is slightly more sensitive than ID +, but shows increased artifacts.

•No difference in performance was observed on the AB 3500 between PP16HS and ID+ during the cooling, denaturation and snap precision, contamination and reproducibility studies.

•No contamination issues were observed during this validation.

•The Anne Arundel County Crime Laboratory will continue to use PP16HS with the AB 3500 for casework analysis.

•SOP will not include denaturation and snap cooling prior to an instrument run.

References

1. Butts, Erica L.R., et al. NIST Validation Studies of the 3500 Genetic Analyzer. Forensic Science International: Genetics Supplement Series 3; 2011:184-185.

2. Butts, Erica L.R., et al. NIST Validation Studies of the 3500 Genetic Analyzer. National Institute of Standards and Technology. Presentation for the Mid-Atlantic Association of Forensic Scientists, May 27, 2011. 3. Butts, Erica L.R., et al. NIST Validation Studies of the 3500 Genetic Analyzer. National Institute of Standards and Technology. Presentation for the 24th Congress of the International Society for Forensic Genetics, Aug. 31, 2011.

. Butts, Erica L.R., et al. NIST Validation Studies of the 3500 Genetic Analyzer. National Institute of Standards and Technology. Presentation for the 22nd International Symposium of Human Identification, Oct. 6, 2011.

5. Qi, Liwei, et al. HID Validation: the 3500 series genetic analyzers. Life Technologies: Applied Biosystems HID University. Presentation. 6. Fryback, Dawn. 3500 Validation. Date of email communication: April 26, 2012.

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